

FineFDR: Fine-grained Taxonomy-specific False Discovery Rates Control in Metaproteomics

Shengze Wang ^[1], Shichao Feng^[1], Chongle Pan^[2], Xuan Guo^[1]

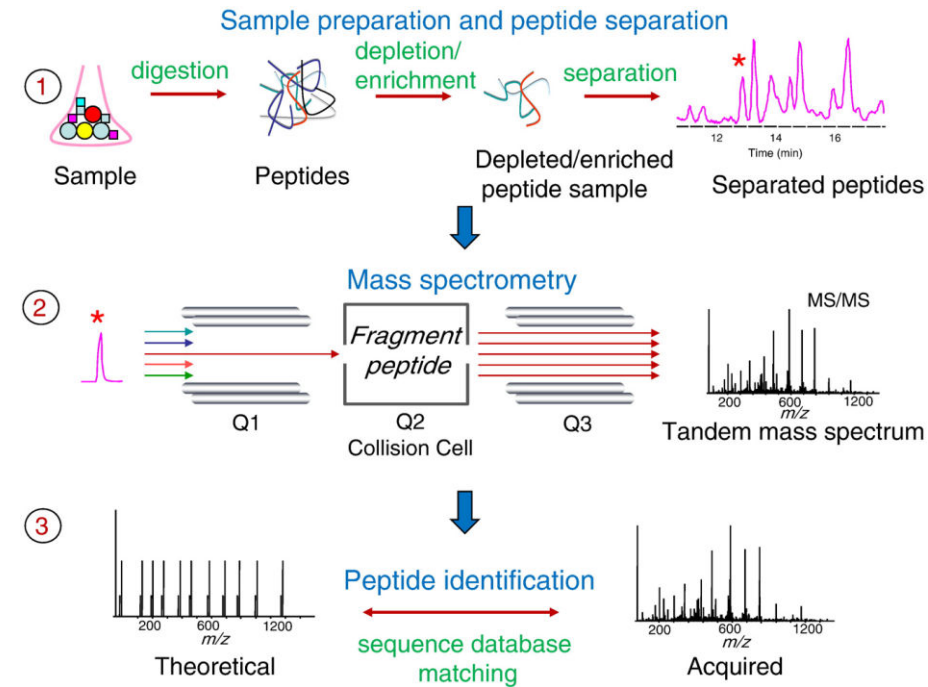
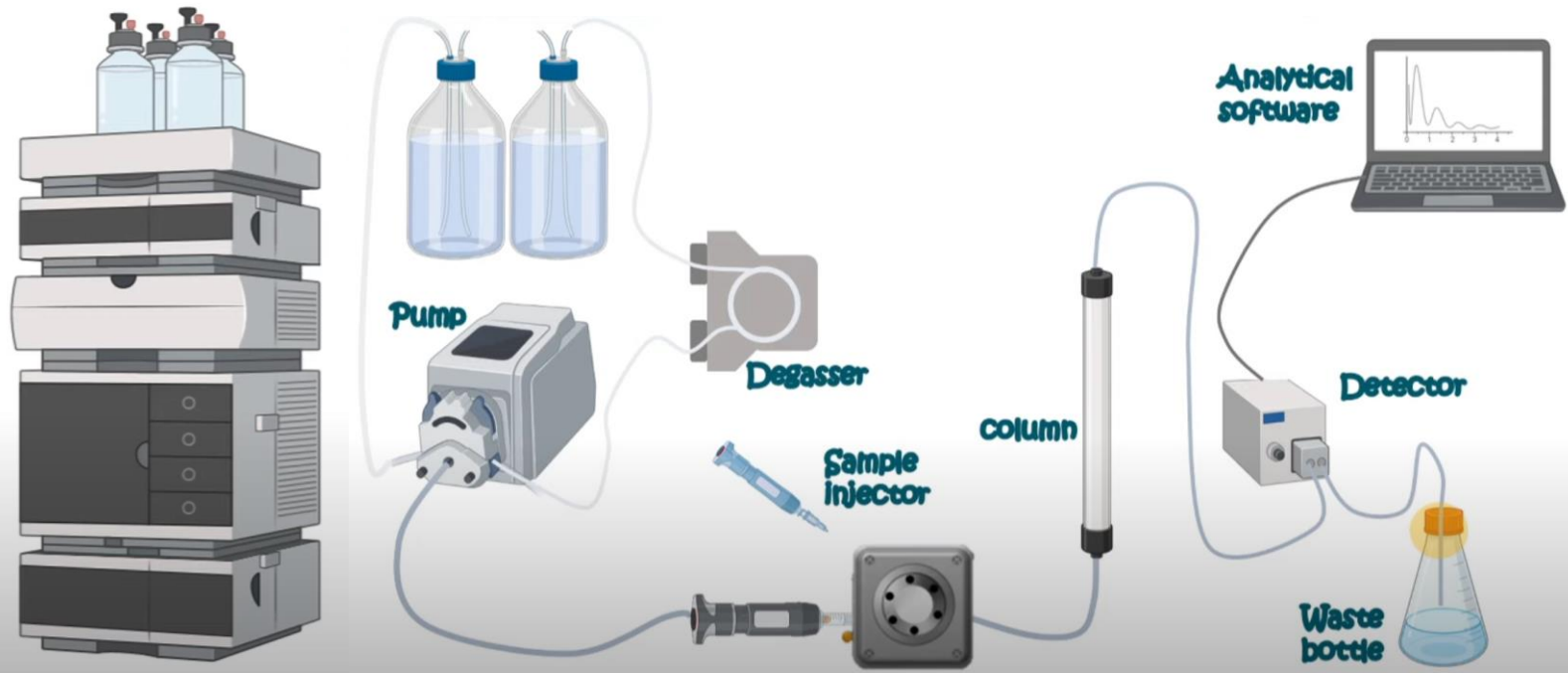
^[1] Department of Computer Science and Engineering, University of North Texas

^[2] School of Computer Science & Department of Microbiology and Plant Biology, University of Oklahoma

1

Current discovery metaproteomics studies

are generally based on high-throughput tandem mass spectrometry (MS/MS) coupled with liquid chromatography (LC). (LC-MS/MS)



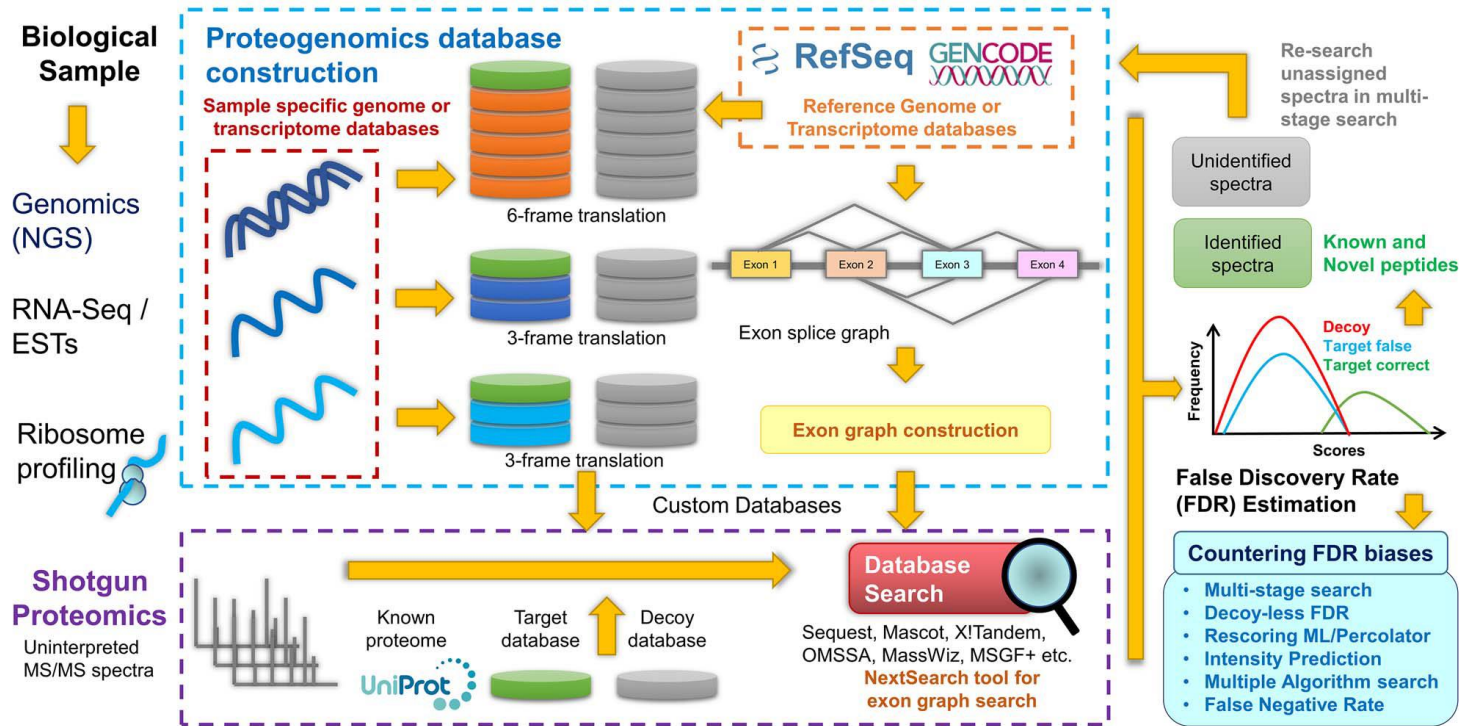
Source: A. biology With arpan, "HPLC | High Performance Liquid Chromatography | Application of HPLC," 16-Sep-2020. [Online]. Available: <https://www.youtube.com/watch?v=Vr5t-cgHHG4>. [Accessed: 23-Nov-2022].

Source: A. I. Nesvizhskii, "A survey of computational methods and error rate estimation procedures for peptide and protein identification in shotgun proteomics," Journal of Proteomics

2

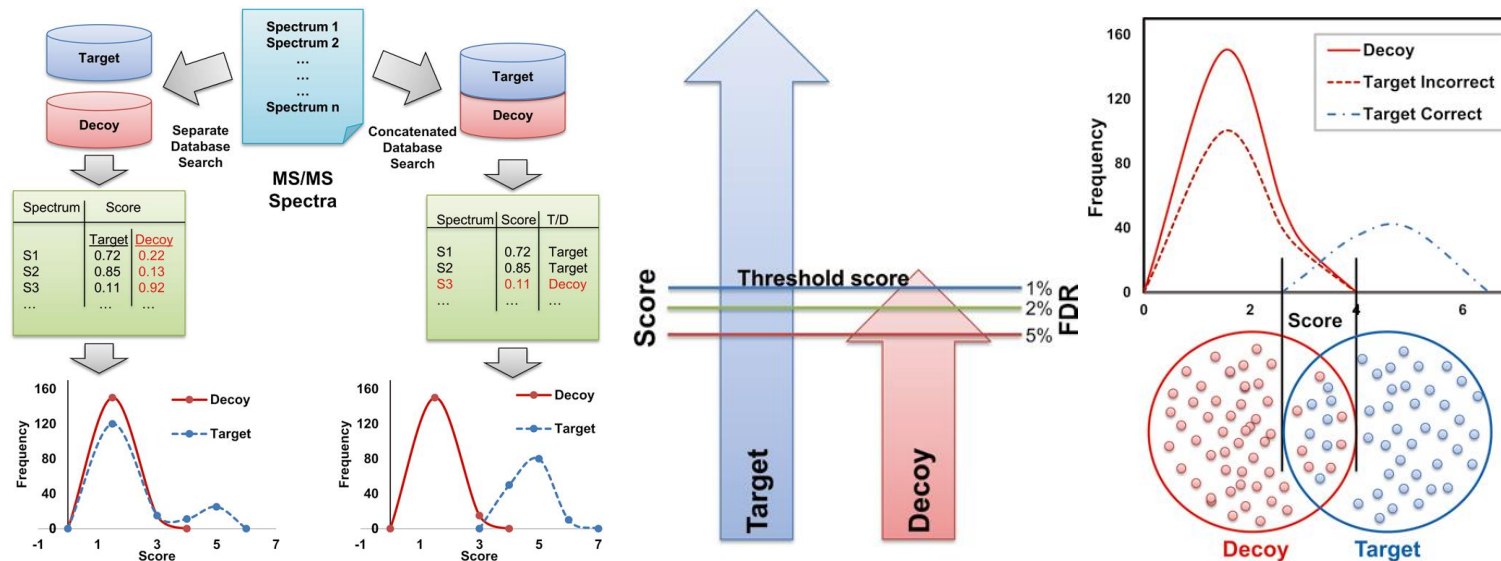
Identifying peptides and proteins from microbiota

involves a procedure of searching mass spectra against a pre-defined protein sequence database.



Source: S. Aggarwal, A. Raj, D. Kumar, D. Dash, and A. K. Yadav, "False discovery rate: the Achilles' heel of proteogenomics," Briefings in Bioinformatics, vol. 23, no. 5. Oxford University Press (OUP), May 09, 2022. DOI: 10.1093/bib/bbac163.

- 3** A major post-analysis step is controlling the **false discovery rate**, i.e., **FDR**, the ratio of false positives to the total number of annotations.
- 4** The current gold standard for FDR estimation is the target-decoy search strategy using p-value or E-value.



$$FDR = \frac{\# Decoys}{\# Targets}$$

Source: S. Aggarwal and A. K. Yadav, "False Discovery Rate Estimation in Proteomics," Methods in Molecular Biology. Springer New York, pp. 119–128, 2016. doi: 10.1007/978-1-4939-3106-4_7.

Reliability of identifications

a. Single-identification level

p-value or E-value

b. Multiple-identification level

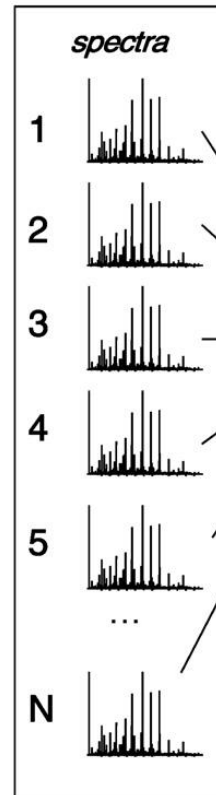
The proportion of incorrect identifications for a group of identifications.



The problem of FDR estimation in multiple hypothesis tests:

Treat all the peptides and proteins equally and overlook that they could have varied probabilities of being identified.

Entire dataset, N spectra



database search

Filtering using target-decoy strategy

Best match for each spectrum

spec	peptide	score	label
1	ISLLDAQSAPLR	4.5	target
2	VVEELCTPEGK	3.9	target
5	GDAVFVIDALNR	3.6	target
3	VNSPMKWVPTPK	1.7	decoy
4	ECDVVSNTIIAEK	1.5	target
...
N	LIHSVFGIGEK	1.1	decoy

(sorted by score)

Apply score threshold S_T
Calculate $N_t(S_T)$ and $N_d(S_T)$:
number of target/decoy PSM with $S \geq S_T$
Estimate FDR $FDR(S_T) = \frac{N_d(S_T)}{N_t(S_T)}$
Select threshold S_T to achieve desired FDR

Source: A. I. Nesvizhskii, "A survey of computational methods and error rate estimation procedures for peptide and protein identification in shotgun proteomics," Journal of Proteomics, vol. 73, no. 11. Elsevier BV, pp. 2092–2123, Oct. 2010.

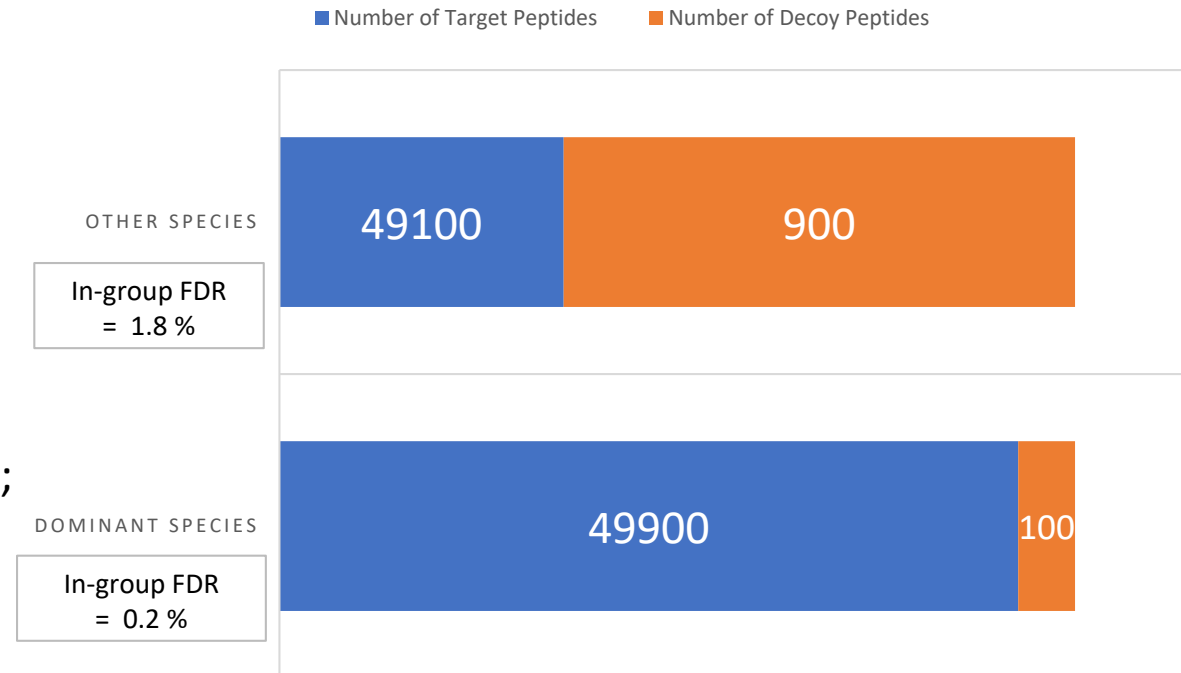
Motivation

The problem of FDR estimation in multiple hypothesis tests:

Treat all the peptides and proteins equally and overlook that they could have varied probabilities of being identified.

In an extreme case, If we have
50,000 identified peptides from a dominant species,
50,000 identified peptides from other species;
FDR level is set to be 1%,
so expected false-positive identification = $100,000 \times 0.01 = 1,000$;
Varied probabilities of being identified:
10% of false-positive were from the dominant species, and the left was from the other species.

ACTUAL IN-GROUP FDR IN THE CASE





Main idea

FineFDR controls the FDR separately for PSMs/peptides/proteins from the different taxonomic units.

Assumption

Peptides and proteins are not equally likely to be measured by LC-MS/MS and identified by search engines due to the varied abundance of microorganisms.

Method: Target-decoy FDR Control

The basic target-decoy strategy augments the "target" protein database with a set of "decoy" protein sequences.

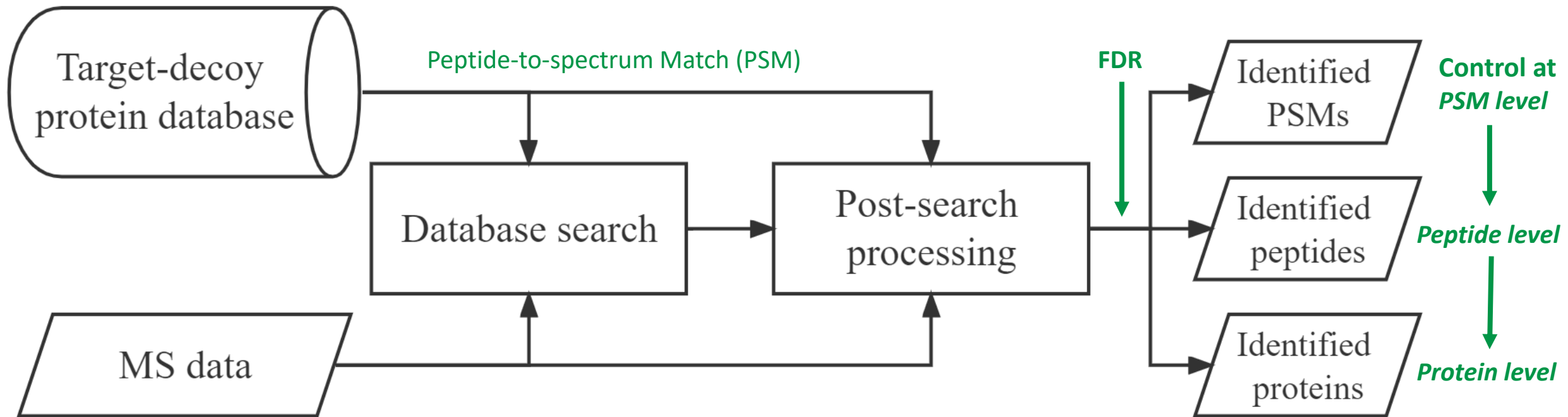


Fig. 1. The basic workflow of the target-decoy search strategy.

Method: Taxonomy Database Construction

Operational Taxonomic Unit (OTU)

- Groups of closely related microorganisms at the genome level
- Basic unit to group PSMs or peptides in FineFDR

Peptide-to-Spectrum Matches (PSM)

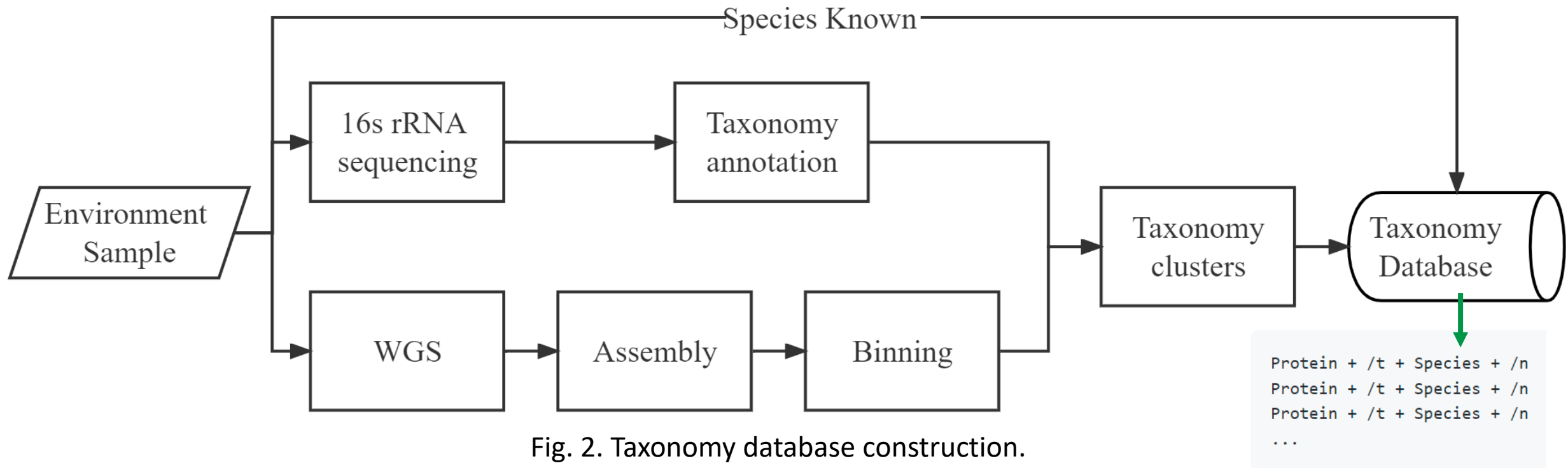


Fig. 2. Taxonomy database construction.

Method: Taxonomy-specific FDR assessment (PSM)

$$\text{In-group FDR}_{OTU\ index\ i} = \frac{\# Decoys \subset OTU\ Cluster_i}{\# Targets \subset OTU\ Cluster_i}, \quad \text{Global FDR}_i = \frac{\# Decoys}{\# Targets}$$

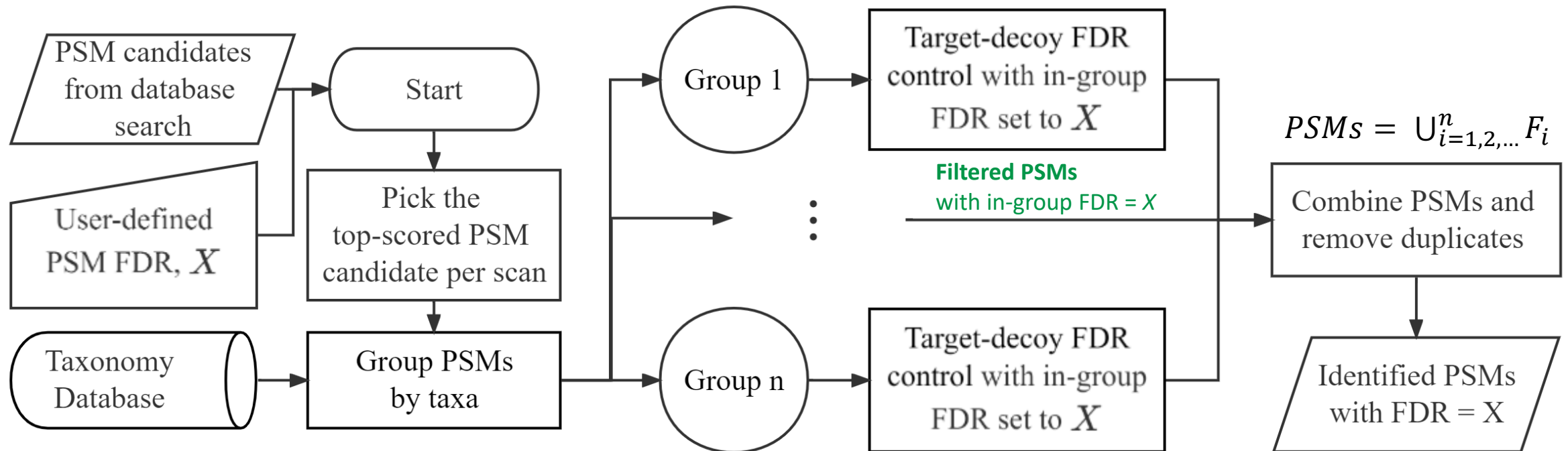


Fig. 3. The framework of taxonomy-specific FDR control at the PSM level.

Method: Taxonomy-specific FDR assessment (Peptide)

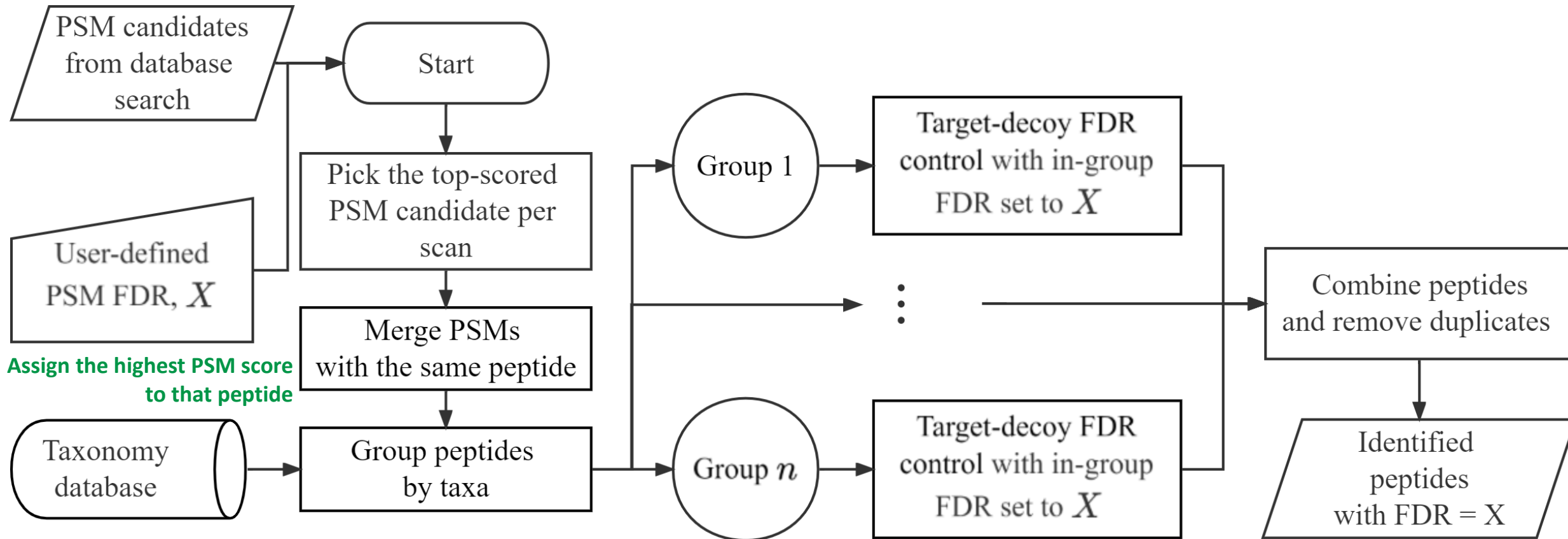


Fig. 4. The framework of taxonomy-specific FDR control at the peptide level.

Method: Taxonomy-specific FDR assessment (Protein)

Because transferring PSM scores to protein scores is not trivial, FineFDR adjusts in-group peptide and protein FDRs dynamically *Until* the global protein FDR is well controlled.

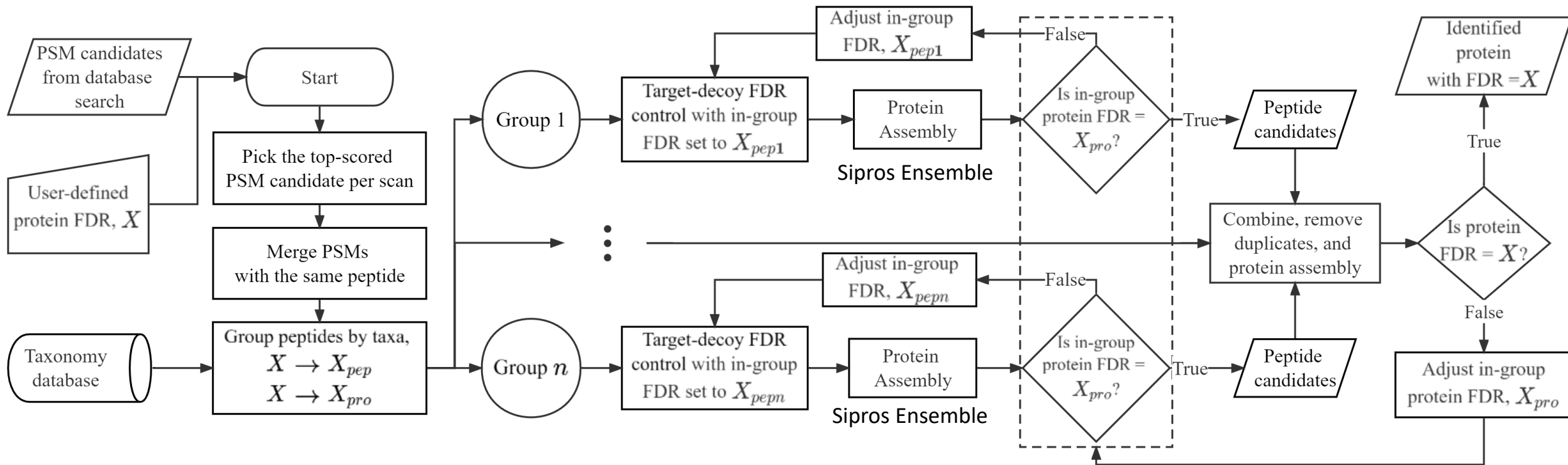


Fig. 5. The framework of taxonomy-specific FDR control at the protein level.

Experiment Design

Search Tool

- Comet (E-value score; Widely-used method)

Filtering Tool

- Percolator (Percolator score; Widely-used method)
- TIDD (TIDD SVM Prob; Recent work)
- Tailor (Tailor score; Recent work)

Data sets

- Simulated data set: Mock microbial "U" (UNEVEN) type community data set with the cell number U1 (PXD006118)
- Real-world data set: Marine 1,2,3 (PXD007587); Soil 1,2,3 (PXD007587); Human Gut (PXD013386)
- Simulated ground-truth data set: GT

TABLE I
THE TOTAL NUMBERS OF MS/MS OF METAPROTEOME DATA SETS

Data Set	Mock U1	Marine1	Marine2	Marine3	Soil1	Soil2	Soil3	Human Gut	GT
# of spectra	390,110	138,682	143,344	127,075	421,606	505,477	367,265	141,811	141,811

Identification Quality

$$\textit{Precision} = \frac{\# \textit{True identifications}}{\# \textit{Identifications}}$$

Ground-truth data set simulation

Search the Human Gut’s MS data against the database consisting of the protein mixture from the Human Gut and Marine protein databases.

“Truth” identifications

The PSMs/peptides/proteins from the Human Gut proteome.

Result

FineFDR achieved higher precision than the baseline methods.

TABLE II
BENCHMARKING OF IDENTIFICATION PERFORMANCE USING THE
GROUND-TRUTH DATA SET

Search + Filter ^a	True	False	Precision
# PSM identifications at FDR 1%			
C	34,902	772	0.978
C w/F	48,585	211	0.996
C + P	39,480	2,869	0.932
C + P w/F	60,303	2,350	0.960
TIDD	38,098	875	0.978
TIDD w/F	50,805	165	0.997
Tailor	31,793	362	0.989
Tailor w/F	51,736	82	0.998
# Peptide identifications at FDR 1%			
C	12,432	296	0.977
C w/F	17,356	132	0.992
C + P	14,200	1,920	0.881
C + P w/F	20,276	1,892	0.915
TIDD	13,447	357	0.974
TIDD w/F	18,286	126	0.993
Tailor	11,313	134	0.988
Tailor w/F	19,003	47	0.998
# Protein identifications at FDR 1%			
C	1,622	106	0.968
C w/F	4,110	37	0.991
C + P	3,588	1,622	0.689
C + P w/F	4,602	1,673	0.733
TIDD	3,274	106	0.972
TIDD w/F	4,434	0	1.000
Tailor	2,454	67	0.973
Tailor w/F	3,492	46	0.987

^a Searching\Filtering algorithms: C, Comet; F, FineFDR; P, Percolator.

Identification Rate



TABLE III
BENCHMARKING OF IDENTIFICATION PERFORMANCE USING EIGHT METAPROTEOMES

Search + Filter ^a	C	C + F	C + P	C + P + F	TIDD	TIDD + F	Tailor	Tailor + F
# PSM identifications at FDR 1%								
Mock U1	125,517	130,541	130,166	130,317	116,238	119,236	129,909	137,726
Marine1	43,125	48,456	52,039	52,932	38,979	44,690	38,670	47,252
Marine2	38,753	43,921	48,569	49,411	33,817	39,883	31,639	40,167
Marine3	46,781	52,312	54,506	55,446	43,684	48,705	43,938	52,847
Soil1	50,374	52,937	55,219	55,298	51,189	52,221	56,162	59,662
Soil2	43,832	46,498	48,823	48,878	48,222	49,692	51,929	55,285
Soil3	57,141	60,612	63,572	63,730	NaN ^b	NaN ^b	59,769	64,643
Human Gut	43,565	46,316	48,108	48,154	48,439	49,315	48,439	49,315
# Peptide identifications at FDR 1%								
Mock U1	47,674	49,136	49,786	49,921	47,260	48,110	47,717	49,808
Marine1	26,960	30,622	35,253	35,837	24,404	28,099	22,491	27,288
Marine2	27,166	31,133	36,902	37,498	23,575	28,110	20,727	26,467
Marine3	30,886	34,589	38,313	38,903	28,913	32,394	26,878	32,410
Soil1	17,050	17,853	19,048	19,525	16,484	16,880	14,620	15,599
Soil2	15,473	16,273	17,311	17,767	14,949	15,459	12,817	13,642
Soil3	16,872	17,761	19,128	19,734	NaN ^b	NaN ^b	13,832	14,791
Human Gut	15,396	16,646	17,527	18,055	16,885	17,686	16,855	17,686
# Protein identifications at FDR 1%								
Mock U1	8,740	8,784	9,135	9,155	8,838	8,865	8,579	8,743
Marine1	8,101	8,599	13,816	14,065	7,579	8,233	6,230	7,052
Marine2	8,677	9,325	15,788	16,065	7,634	8,441	6,098	7,112
Marine3	9,172	9,832	13,993	14,328	8,372	9,167	7,567	8,506
Soil1	4,823	5,031	5,204	5,432	4,790	4,937	4,136	4,387
Soil2	5,090	5,294	5,650	5,789	5,012	5,014	4,082	4,360
Soil3	5,012	5,188	5,418	5,595	NaN ^b	NaN ^b	4,131	4,407
Human Gut	3,779	3,956	4,140	4,360	4,064	4,186	4,064	4,186

^a Searching/Filtering algorithms: C, Comet; F, FineFDR; P, Percolator.

^b Unable to generate any results due to program error exceptions

Identification Rate
Number of PSMs,
peptides, and proteins
filtered at 1% FDR

Result
For the methods
adding FineFDR, they
achieved more
identifications than
the baseline methods
without FineFDR.

Computational time

Test Platform

A regular desktop with an 8-Core 4.0 GHz CPU, 32GB 3200 MHz RAM, and NVMe 3.0 SSD.

FineFDR is implemented with Python 3.9.

On average, FineFDR requires 2 GB of memory to load data.

Table S1. The computational time of FineFDR

Data sets	Average time cost on three runs (minutes)
Mock U1	17
Marine Community	36
Soil Community	31
Human Gut Community	25



Discussion

1

The baseline method and its combination with FineFDR shared over 95% identical PSMs, peptides, and proteins in the results. FineFDR made more method-specific discoveries than the baseline method.

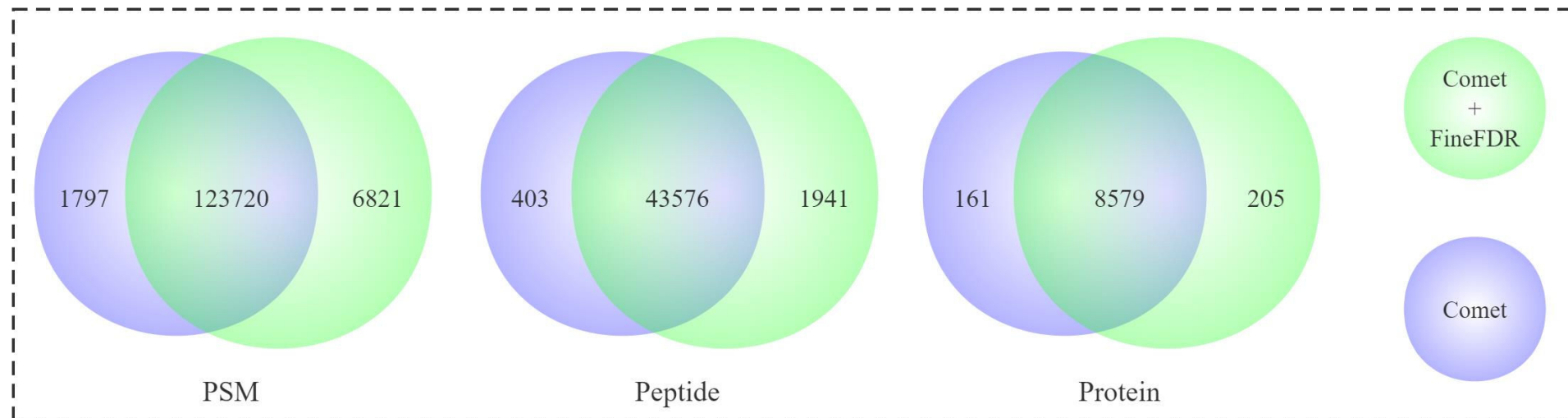


Fig. 6. The identified result overlap between the baseline method comet and its combination with FineFDR for the Mock U1

Discussion

2

FineFDR improved the identification rates across most species.

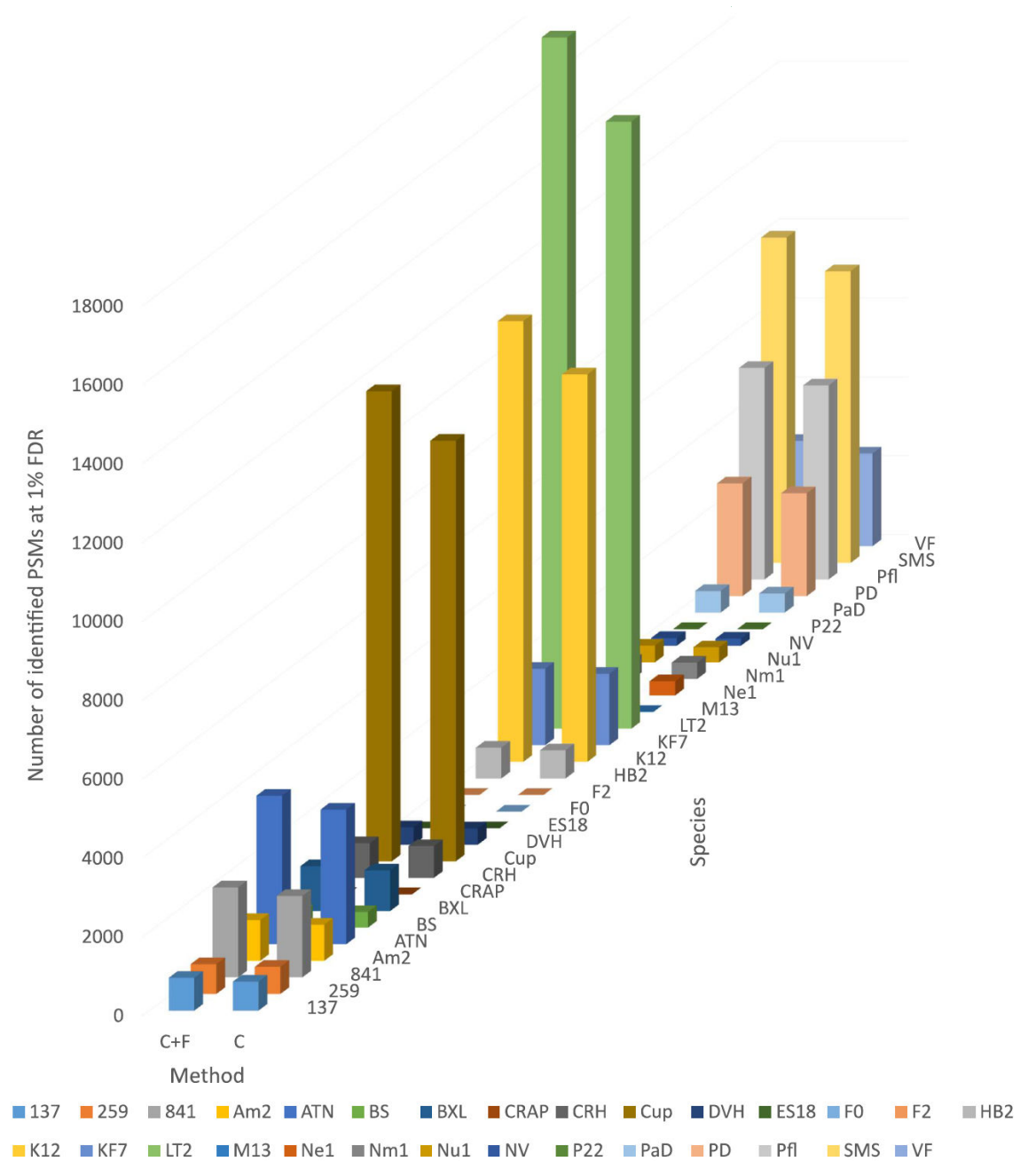


Fig. S1. PSM identification improvements by species for the Mock U1

Discussion

3

FineFDR shows the power to promote the percentage of target PSM candidates in a group

Table S2. Number of PSMs in Marine 1 before applying FineFDR

Data set	Target	Decoy	Target/(Target + Decoy)
Marine 1	93906	39851	0.70206419

The percentage of target PSM candidates in a random group without efficient grouping would be close to that in the original method without grouping.

Table S3. Number of PSMs by species with duplicate PSMs across the groups in Marine 1

Species	Target	Decoy	Target/(Target + Decoy)
output.marine.1.fa.pin	36	11	0.765957447
output.marine.10.fa.pin	293	66	0.816155989
output.marine.100.fa.pin	869	67	0.928418803
output.marine.101.fa.pin	775	87	0.899071926
output.marine.102.fa.pin	906	106	0.895256917
output.marine.103.fa.pin	538	83	0.866344605
output.marine.104.fa.pin	388	4	0.989795918
.....			
.....			
output.marine.9.fa.pin	1291	107	0.923462089
output.marine.90.fa.pin	525	74	0.876460768
output.marine.91.fa.pin	119	57	0.676136364
output.marine.92.fa.pin	658	83	0.887989204
output.marine.93.fa.pin	609	54	0.918552036
output.marine.94.fa.pin	688	54	0.92722372
output.marine.95.fa.pin	535	65	0.891666667
output.marine.96.fa.pin	2399	130	0.948596283
output.marine.97.fa.pin	330	61	0.84398977
output.marine.98.fa.pin	1311	154	0.894880546
output.marine.99.fa.pin	1097	39	0.965669014
Unknown.pin	73879	31507	0.701032395

Conclusion



Contribution

- A novel FDR estimation framework, called FineFDR, was proposed for metaproteomics.
- FineFDR controls the FDR separately for PSMs/peptides/proteins from the different taxonomic units.
- FineFDR achieved higher precision and more PSM, peptide, and protein identifications.
- FineFDR is freely available under the GNU GPL license at <https://github.com/Biocomputing-Research-Group/FDR>.

Future Work

- FineFDR will support more search engines and post-search tools in future releases.
- Beyond Taxonomy-specific FDR control, we are investigating more techniques to mitigate the FDR estimation bias in metaproteomics.

Thank you for your time!

CORRESPONDENCE

xuan.guo@unt.edu

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